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RECEIVED 18 August 2023 ACCEPTED 02 October 2023 PUBLISHED 10 October 2023

CITATION

Glaubitz J, Asgarbeik S, Lange R, Mazloum H, Elsheikh H, Weiss FU and Sendler M (2023) Immune response mechanisms in acute and chronic pancreatitis: strategies for therapeutic intervention. *Front. Immunol.* 14:1279539. doi: 10.3389/fimmu.2023.1279539

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Immune response mechanisms in acute and chronic pancreatitis: strategies for therapeutic intervention

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Acute pancreatitis (AP) is one of the most common inflammatory diseases of the gastrointestinal tract and a steady rising diagnosis for inpatient hospitalization. About one in four patients, who experience an episode of AP, will develop chronic pancreatitis (CP) over time. While the initiating causes of pancreatitis can be complex, they consistently elicit an immune response that significantly determines the severity and course of the disease. Overall, AP is associated with a significant mortality rate of 1-5%, which is caused by either an excessive pro-inflammation, or a strong compensatory inhibition of bacterial defense mechanisms which lead to a severe necrotizing form of pancreatitis. At the time-point of hospitalization the already initiated immune response is the only promising common therapeutic target to treat or prevent a severe disease course. However, the complexity of the immune response requires finebalanced therapeutic intervention which in addition is limited by the fact that a significant proportion of patients is in danger of development or progress to recurrent and chronic disease. Based on the recent literature we survey the disease-relevant immune mechanisms and evaluate appropriate and promising therapeutic targets for the treatment of acute and chronic pancreatitis.

KEYWORDS

acute pancreatitis, chronic pancreatitis, immune response, fibrosis, therapy

1 Introduction

Acute pancreatitis is an inflammatory disease of the pancreas which constitutes, with an annual incidence rate of ~34 per 100,000 people, one of the most common diseases of the gastrointestinal tract (1). A significant proportion of patients which experience an episode of acute pancreatitis (about 20-30%) develop a chronic form of the disease (2). Chronic pancreatitis (CP) is defined as a fibroinflammatory disorder of the pancreas, which is mostly characterized by recurrent episodes of AP, and results in a progressive loss of exocrine and endocrine tissue (3). Acute and chronic pancreatitis are clinically defined

10.3389/fimmu.2023.1279539

pictures of a common underlying disease, where immune response mechanisms orchestrate the repair, resolution or fibrotic replacement of acinar cell damage. Acute pancreatitis is defined by a self-digestion of pancreatic tissue by its own proteases (4). The most important etiologic factors include gallstones, alcohol abuse, genetic risk, trauma, metabolic disorders, drugs, infections, and autoimmune causes. The intra-acinar activation of trypsinogen to trypsin has been hypothesized as the initial triggering event in the disease process (5), which ultimately leads to necrotic cell death of acinar cells (6). In recent years, however, it has become increasingly clear that also trypsin-independent cellular mechanisms can initiate cell damage and are thus causative for the development of pancreatitis (6). Genetic functional analyses of sequence variants identified in patients with chronic pancreatitis (CP) unravelled the cellular pathomechanisms which are involved in acinar cell damage. In addition to mutations related to trypsin activity (7-9), misfolding mutations were detected that cause increased endoplasmic reticulum (ER) stress (10-12). Mutations in the cystic fibrosis transmembrane conductance regulator (CFTR) are also related with an increased CP risk and were found to associate with decreased pancreatic secretion (13-15). Whereas these roles of genetic risk factors are relatively well studied, the cellular pathomechanism behind ethanol-toxic pancreatitis is largely not understood. Another major cause is an obstruction of the pancreatic duct by bile stones, but the cellular mechanisms here have also not been fully elucidated. It is assumed that an increased pressure buildup in the pancreatic duct triggers the opening of pressure-sensitive Ca²⁺ channels and results in cytoplasmatic Ca²⁺ overload leading to intracellular protease activation (16).

Independent from etiology and triggering factors of an acute episode of pancreatitis we observe an immediate response of the immune system to local cell damage which ultimately defines the further course of the disease. Acinar cells activate the transcription factor NF κ B in parallel to the Ca²⁺ signal and initiate the expression of cytokines and chemokines which start the local immune response. They mobilize cells of the innate immune system such as monocytes and neutrophils from the bone marrow, and recruit them into the damaged tissue area of the pancreas (17). Tissue resident macrophages are among the first cells of the innate immune system which recognize damage associated molecular pattern (DAMPs) and cytokines released from acinar cells and enhance the initial local immune response. After the onset of the local response more immune cells start to migrate into the organ where they get activated by DAMPs (18, 19). Activated leukocytes in turn stimulate the pro-inflammatory response as well as the local damage by releasing further cytokines, such as TNF- α (20) or reactive oxygen species (ROS) (21). In the later stage of the disease, the inflammatory response changes from a pro-inflammatory toward an anti-inflammatory phenotype which is a prerequisite for wound healing and fibrogenesis (22, 23). Repeated acute episodes frequently progress to chronic pancreatitis which associates with a continuous fibrotic replacement of exocrine and endocrine tissue.

Whereas triggering causes of pancreatitis can vary, the immune response to cell damage is identical and therefore may be best suited for clinical intervention. Due to the complex and dynamic regulation of the immune response in the disease course, any intervention needs to be well balanced and done with caution. In the following we will review in more detail the involvement and therapeutic potential of important signaling pathways and regulation mechanisms in acute and chronic pancreatitis. About 20% of acute pancreatitis patients develop a severe disease course with systemic complications, peripheral organ failure or the infection of pancreatic necrosis by commensal intestinal bacteria (24). Severe disease courses are caused by derailed or unbalanced immune responses and are associated with high mortality and morbidity. The treatment of severe acute pancreatitis is a clinical challenge because the therapy needs to target complex immune regulation mechanisms, which mostly have been investigated in animal models. The translation of therapeutic strategies into the clinic is promising, but also needs to consider existing differences of mouse and human immunity.

2 Acute pancreatitis

2.1 NFκB initiates the inflammatory response

The onset of acute pancreatitis is characterized by an induction of cell damage in the exocrine pancreas, which ultimately results in acinar cell death. Caused by intracellular Ca²⁺ signaling and the cell damage, the transcription factor NFKB becomes activated (17). NFκB is found in almost all cell types and plays a key role in regulating inflammatory responses to stimuli such as stress, cytokines, free radicals, ultraviolet irradiation, danger associated molecular patterns and bacterial or viral antigens. NF-KB controls inflammation not only directly by increasing the production of inflammatory cytokines, chemokines and adhesion molecules, but also by regulating the cell proliferation, apoptosis, morphogenesis and differentiation. Therefore, NFkB could be a possible therapeutic target to control the onset of the immune response and to mitigate the disease progression at an early stage. There are several clinically well-studied and approved inhibitors of the NFKB pathway available, such as aspirin or cyclosporine A (25). Experimental work has shown that NFkB is involved not only in the regulation of immune-related genes, but also in an anti-stress response (26). In an experimental pancreatitis model in mice, the acinar cell specific knockout of RelA, coding for the p65 subunit of NFKB, significantly increased disease severity (27). Likewise, the acinar cell-specific knockout of IKBa, an intracellular inhibitor of NFKB phosphorylation, was shown to increase NFkB activity and resulted in a milder disease course (26). The effect is different when we look at NFKB activation in leukocytes. In pancreatic macrophages, or bone marrow-derived macrophages (BMDMs), NFκB activation stimulates the release of cytokines and chemokines (18), which drives the systemic hyperinflammation and generates a "cytokine storm". Apparently, signaling pathways can have cell specific effects, which limits their eligibility for a broad pharmacological intervention. Instead, both, the molecular target, as well as the target cell must be carefully selected and specifically addressed analogous to a targeted cell-specific tumor therapy.

2.2 The local immune response

Based on observations from the mouse model of caeruleininduced pancreatitis, the local damage scenario in the pancreas has two temporally separated peaks (4, 5, 28). The first peak comprises the acinar cell damage (5, 29), which is caused by intracellular protease activation or massive ER stress and occurs within the first hour after induction of the disease. The second peak of damage is mainly triggered by infiltrating leukocytes (18, 20-22, 30) and is significantly more severe as it can develop into systemic complications (Figure 1). Animal studies have shown that cells of the innate immune response, such as neutrophils and macrophages rapidly migrate into the pancreas after the onset of the disease (20, 21). In contrast, the cells of the adaptive immune system, such as CD4⁺ and CD8⁺ T-cells migrate significantly later into the inflamed pancreas (31, 32). They have a crucial role in the fibroinflammatory process during chronic pancreatitis (33-35). In the first step damaged acinar cells initiate an immune response via NFKB activation and the release of cytokines and chemokines which recruit leukocytes to the side of inflammation. Chemokines such as MCP-1, RANTES, MIG-1, IP-10, and IL8 are significantly elevated in the serum of patients and attract cells to the pancreas (36-38). In parallel to the chemokine mediated cell recruitment, necrotic acinar cells trigger via DAMPs the activation of tissue resident macrophages and initiate the release of cytokines (39). Tissue resident macrophages represent a significant population of quiescent immune cells in the exocrine and endocrine pancreas which react immediately on appearing damage signals (39). These activated local macrophages start a self-enhancing immune response by secreting pro-inflammatory cytokines and chemokines to recruit more immune cells to the side of inflammation (18, 22, 40). However, it is important to note that tissue resident macrophages are also essential to induce cell regeneration and therefore are necessary to minimize the acute damage during pancreatitis. Recent work has shown that especially the induction of fibrosis is a critical protective factor for the survival



FIGURE 1

Schematic illustration of the temporal development of pancreatic damage and the local immune response during acute pancreatitis. In three different phases of the disease course, it is shown how cells of the innate and adaptive immune system are involved in the processes of acinar necrosis, the clearance of damaged tissue and the regeneration of pancreatic tissue or it's fibrotic replacement. Tissue resident macrophages (grey) are the first cells which become activated and differentiate into classical or alternatively activated macrophages, similar to the monocyte derived macrophages. The AP-induced immune response showed similarities to classical wound healing reactions which results in fibrosis and scar formation. Multiple episodes of AP thus result in increased organ fibrosis which characterises the clinical picture of CP.

of severe acute pancreatitis. Depletion of tissue resident macrophages by CSF1 Antibody + clodronate containing liposomes significantly reduced the survival of animals and impaired the tissue regeneration after acute pancreatitis (41). It seems that macrophages have opposing roles in pancreatitis, therefore, a therapeutic treatment involving macrophages must be applied with caution as it could be a double-edged sword.

Neutrophils are among the first immune cells which are recruited to the damaged organ. In contrast to macrophages, they are not tissue resident cells, but extravasate from the vascular system. Antibody-mediated depletion of neutrophils in animal models of acute pancreatitis significantly ameliorated the disease course (20, 21, 38). A crucial role in this process plays the excessive release of reactive oxygen species (ROS) via oxidative burst, a defense mechanism against pathogenic bacteria (42). The excessive release of ROS causes additional damage of pancreatic tissue (21) and also damages the vascular system, which in turn stimulates the migration of more immune cells (42). Another defense mechanism of neutrophils against pathogens is the formation of net-like chromatin fibers decorated with antimicrobial proteins, which trap and kill bacteria to prevent further spreading in the organism (43). NETs (Neutrophile extracellular traps) are also formed during acute pancreatitis, and animal studies have shown that these NETs increase the local damage and therefore could be a potential therapeutic target (44, 45). Leppkes et al. analyzed the underlying pathomechanism and showed that NETs are involved in stone formation and contribute to pancreatic duct obstruction (46). They proclaim that one of the main triggers for NET formation is the pro-inflammatory cytokine IL-17A, which is released by a subset of pro-inflammatory T helper cells (Th17 cells). T-cell infiltration into the pancreas occurs in the middle or later phase of the disease, but the activation of T-cells can be observed in the lymphoid tissue (40) or the small intestine (47,48), where they play a crucial role for the intestinal barrier function. Beside IL-17A also pathogens, PAMPs and DAMPs can induce NET formation in neutrophils (49)however, AP is a primarily sterile inflammation, so it seems unlikely that pathogens or PAMPs cause NET formation in AP Mechanical stimuli such as microcrystals can also trigger netosis and have been found to play a crucial role in the formation of gallstones from biliary crystals (50). They could also be a direct cause of AP, independent of Th17-cells. IL-17A acts directly on neutrophils and recruits them to the side of inflammation (51). A limited T-cell infiltration in the early phase of the disease may provoke also a limited neutrophil migration (18). Neutrophils are rather out of place during pancreatitis, because in the beginning, pancreatitis is a primarily sterile inflammation and neutrophils are mainly involved in bacterial clearance. Still, their main function is to trap and kill bacteria via NETs and ROS release, and both functions increase the local organ damage during pancreatitis (21, 46) (Figure 1).

In contrast to neutrophils, monocytes react more flexible to the local circumstances. Chemokines like CCL2 (MCP-1) are involved in the mobilization and recruitment of monocytes. CCL2-binding to its receptor CCR2 recruits mainly pro-inflammatory Ly6C^{hi} monocytes, but also neutrophils and T-cells to the site of inflammation (52). While circulating monocytes migrate to the

pancreas, tissue-resident macrophages are the first immune cells that react immediately to the pancreatic damage. A therapeutic blockade of the CCR2 pathway diminishes the migration of monocytes and reduces the local as well as the systemic damage (38, 53, 54). Notably, this blockade does not significantly affect the local macrophage response, as tissue resident macrophages start to proliferate and are able to substitute the monocyte derived macrophage response (38). Monocyte derived macrophages enter the pancreas almost as fast as neutrophils (18, 20, 38), but in contrast to neutrophils, macrophages are, in addition to a bacterial clearance also responsible for the removal of cellular debris and dead cells. In acute pancreatitis macrophages phagocytose the remains of damaged or dead acinar cells (18, 55). Crucial for the clearance process is the nature of the damage (56, 57). Apoptotic cell bodies are immunologic silent, as they do not induce a proinflammatory response (57). Necrotic cells are different, because they release intracellular organelles and proteins which act as DAMPs and induce a pro-inflammatory phenotype of macrophages similar to the response to pathogen associated molecular pattern (PAMPs) (56). In pancreatitis, a switch from necrotic cell death to apoptotic cell death can ameliorate the disease course (58), but acinar cell death is mainly due to necroptotic cell death (59). Necroptosis is a form of regulated necrosis, which is induced by TNF- α via the TNF receptor signaling pathway (60). Like in necrosis it leads to the release of DAMPs, which induce the pro-inflammatory macrophage phenotype and cause secretion of cytokines TNF- α , TRAIL, IL-6, IL-1 β , IL-18 and chemokines such as CCL2, CCL5, CXCL3 and IP-10 (18, 22, 40). Consequently, this initiates a self-sustaining cycle of TNF-\alpha-secreting macrophages which in turn induce necroptosis of acinar cells and the release of more DAMPs to trigger the proinflammatory reaction. Proinflammatory macrophages secrete the majority of cytokines which fuel the upcoming cytokine storm and cause the manifestation of the inflammation (6, 18, 20, 22, 30, 40). The extent of the initial acinar cell damage seems critical in shaping the macrophage response, which in turn enhances the local damage and triggers the development of a cytokine storm and systemic complications. As macrophages are essential for the clearance of the damaged cells in the organ, they cannot be easily targeted in a therapeutic approach (55). Instead, the released cytokines, which play a major role in controlling the systemic immune response, could be addressed (Figure 1).

2.3 Cytokine storm

TNF- α looks like a promising target to interrupt this vicious cycle, depletion of TNF- α reduced the pancreatic damage via inhibiting necroptosis and diminished the pro-inflammatory response (60). A depleting antibody (Infliximab) is already in clinical use for the treatment of rheumatoid arthritis (61), and an ongoing study currently investigates the therapeutic potential of infliximab in patients with acute pancreatitis (NCT03684278). Previous studies on patients suffering from sepsis or Morbus Crohn showed an increased risk of secondary infections, and the mortality risk was significantly higher in sepsis patients (62, 63). An

extended blockage of pro-inflammatory signaling could strengthen the anti-inflammatory response and increase the risk of secondary infections, which are known to be a serious risk also during severe acute pancreatitis (24). The current study will show if anti-TNF- α treatment is effective and safe in pancreatitis patients.

In addition to TNF- α , activated macrophages secrete also IL-6, which is another well-known pro-inflammatory cytokine (64). In acute pancreatitis, increased serum levels of IL-6 correlate with disease severity (40, 65). In addition to an activation of the classical IL-6 signaling pathway through the IL-6 receptor/gp130 complex, IL-6 can also act via the IL-6 trans-signaling pathway by interaction with the soluble IL-6 receptor on cells that do not express the classical IL-6 receptor, but only gp-130 (66).. Trans-signaling can have dramatic effects on various cell types. During pancreatitis, IL-6 trans-signaling causes lung damage (67) and defines the systemic disease severity. ADAM17 cleaves the membrane bound IL-6R to its soluble form sIL-6R, which is essential for the IL-6 trans signaling and enables the binding to the gp130-receptor pathway in the absence of membranous IL-6R. The inhibition of ADAM17 results in decreased IL-6 trans signaling and was shown to ameliorate the disease severity of acute pancreatitis (68). Tocilizumab, an antibody which blocks the classical IL-6 pathway by binding to the IL-6 receptor is approved for the treatment of rheumatoid arthritis (69) and was also tested in an animal model for the treatment of acute pancreatitis with promising results (70).

Another proinflammatory cytokine released by macrophages in the course of acute pancreatitis is Interleukin 1 β (71). Pancreasspecific overexpression of IL-1ß results in a form of chronic pancreatitis and causes a complete disintegration of the organ (72). IL-1 β is mainly released by classically activated macrophages. The secretion of IL-1 β is rather complex and requires activation by two independent signals: 1) the induced expression of inflammasome components and of pro-IL-1B via the transcription factor NF κ B, and 2) the initiation of the inflammasome complex formation, consisting of NLRP3, ASC and caspase 1. Only this inflammasome complex is capable of processing pro-IL-1ß by caspase 1 to its mature form. After phagocytosis of necrotic cell debris (18), macrophages express and activate a cytosolic inflammasome complex, which starts the proteolytic maturation of pro-IL-1 β and pro-IL-18 (18, 19, 40). Both cytokines are released from the cytoplasm into the extracellular space by the formation of the gasdermin-D pore complex. Cleavage of gasdermin D by caspase 1 is an essential prerequisite in the pore formation and leads to pyroptotic cell death of the macrophage. Recent data have shown that IL-1ß maturation by the inflammasome complex is a key driver of pro-inflammation, pancreatic tissue damage (71) and systemic inflammation (40, 73). Inflammasome related pathways are accessible by multiple treatment options. The NLRP3 inhibitor MCC950 inhibits the inflammasome complex formation (74), and was shown to ameliorate the disease severity in an animal model of acute pancreatitis (40). Also, treatment with lactate (73, 75) or butyrate (76), which are both known to supress inflammasome activation, are similar effective. The inhibition of the IL-1 β signaling pathway is another treatment option (77). The IL1 receptor antagonist Anakinra, the soluble decoy receptor rilonacept and the neutralizing antibody canakinumab have been tested in multiple diseases like recurrent pericarditis (78) and rheumatoid arthritis (79) or autoinflammatory diseases such as the familial Mediterranean fever (80). Beside these chronic diseases, the IL1 receptor antagonist Anakinra showed beneficial effects also in severe acute conditions like in sepsis patients with features of the macrophage activation syndrome (81). However, not all patients benefited from the treatment, especially when the immunosuppression was too intense (82). The immune mechanisms of severe acute pancreatitis and sepsis are similar, so the therapeutic treatment of the IL1 pathway could be promising, but no clinical studies with AP patients have been performed to date. One single study in a rat model of acute pancreatitis showed that the application of anakinra had a beneficial effect on disease severity (83). Clinical trials are necessary to further evaluate the therapeutical potential of these drugs.

Besides pro-inflammatory cytokines also the anti-inflammatory cytokine Interleukin 10 is increased in serum of patients with AP (65, 84). IL-10 has immunosuppressive functions and inhibits the secretion of pro-inflammatory cytokines via the induction of SOCS3, which would make a stabilized IL-10 protein derivate a good candidate for a clinical treatment (85). IL-10 can reduce the severity of acute pancreatitis in animal models (86, 87), furthermore, it has been shown that the preventive administration of IL-10 can reduce the risk of post-ERCP pancreatitis (88). Notably, IL-10 treatment also comes with a risk; while it can help to control the SIRS, excessive suppression of the immune response can lead to CARS or even immune paralysis. In the course of severe AP we observe a particular and excessive activation of regulatory T cells, which act immunosuppressive via the release of IL-10 (40, 65) and are responsible for the translocation of commensal bacteria from the intestine (47); additional administration of IL-10 could enhance this effect. The induction of a hypo-inflammation can facilitate the translocation of enteral bacteria to pancreatic necrosis and is associated with high mortality, as infected necrosis is one of the highest risk factors for severe disease and fatal outcome of acute pancreatitis (24).

2.4 SIRS/CARS – balance

Development of a cytokine storm, meaning the uncontrolled and excessive release of pro-inflammatory cytokines, can cause serious complications like multisystem organ failure and death (89). Parallel to the massive release of pro-inflammatory cytokines the levels of anti-inflammatory cytokines like IL-10 also rise in a severity dependent manner (40, 65) as it is observed in sepsis (90). The reason behind is an insidious immunological counter-regulation, the so called compensatory anti-inflammatory response syndrome (CARS), a systemic deactivation of the immune system tasked with restoring homeostasis from an inflammatory state. In the past, it was assumed that CARS builds up as a consequence of the excessive proinflammation of SIRS. The increasing auto-immunosuppression in CARS in the worst case, can lead to immune paralysis, which is also associated with serious complications such as infection of pancreatic necrosis by commensal intestinal bacteria (Figure 2A). Pro-

inflammatory mediators such as IL-6, IL-12 or IL-1b which are released during the cytokine storm suppress an anti-inflammatory response (40, 91, 92), and thus further the pro-inflammation. On the other hand, anti-inflammatory mediators such as IL-10 suppress the pro-inflammatory response and enhance the type 2 immune response via the expansion of Tregs, M2 macrophages and Th2 cells (40, 47, 93). Immune response mechanism need to keep a balance between pro- and anti-inflammatory mediators to prevent life-threatening complications during severe AP (Figure 2). Recent studies on sepsis or severe trauma from burn injuries suggest that CARS already develops in parallel with SIRS (91, 92, 94, 95). Matching with the cytokine storm which originates from the local macrophages (18, 19, 22), a systemic counter-regulation is observed (40, 65) (Figure 2B). Early in the disease course an increase of regulatory T-cells (47), Th2 cells (40) and myeloid derived suppressor cells (MDSCs) (65) can be seen and IL-10 levels start to rise (40, 65). Whereas SIRS has clear diagnostic criteria (96), CARS is difficult to evaluate in patients. Lymphocyte dysfunction or increased apoptosis cannot be directly measured for the clinical diagnosis (97), so they often occur unnoticed. Further, increased serum levels of IL-10 are found in patients with severe traumata (92, 98) as well as in patients with severe acute pancreatitis (40, 65). Experimental data show that IL-10 can be protective and is able to reduce the severity of pancreatitis by limiting pro-inflammation (86). A critical limit, where proinflammation switches to immunosuppression is hard to define. The fact that IL-10 increases in parallel with pro-inflammation and that there are no reliable diagnostic marker to detect hypoinflammation at an early stage makes the prediction of severe disease courses very difficult and unreliable. Leukopenia and decreasing pro-inflammation are clear signs of an ongoing CARS, but may, at the timepoint of detection, be too advanced to prevent immune paralysis and the development of secondary infections (24). Improved early diagnosis of CARS is essential for an effective treatment.

The fact that pro-inflammatory and anti-inflammatory mechanisms are active at the same time, suggests that the balancing of the immune response is crucial for the outcome (91, 94, 99). Most therapeutic agents are directed against the excessive



FIGURE 2

Development of SIRS/CARS during acute pancreatitis. (A) Illustrates the temporal correlation of SIRS and CARS with the disease progression. While SIRS frequently leads to organ complications and associates with early mortality, infected necrosis frequently occurs during CARS and causes late mortality. Excessive pro-inflammation suppresses the anti-inflammation and results in an immunological disbalance which is associated with serious complications and an increased mortality (dotted lines). However, excessive anti-inflammation also results in a suppressed ability to respond to bacterial infection and leads to colonization of necrosis by commensal intestinal bacteria, which is also associated with a severe disease course and increased mortality (dotted lines). (B) Recent data suggest a simultaneous induction of pro- and anti-inflammatory responses, which keep the balance during the disease progression. (C) An immunotherapeutic approach needs to preserve this balanced immune response and prevent a shift to either SIRS or CARS. Solid lines mark the immune dysbalance and disease severity of surviving patients, whereas dotted lines indicate a lethal course of the disease.

pro-inflammation in order to mitigate SIRS and its associated complications. These approaches are unfortunately also effective in increasing the counter-regulation (CARS) and thus promoted secondary infections. Similarly, anti-inflammatory therapies would increase the pro-inflammatory response and further advance the SIRS (90). A critical factor in the counter regulation of SIRS is a stringent time control, meaning rapid start and rapid cessation of the treatment. Due to the long-lived nature of neutralizing antibodies such as infliximab (anti-TNF- α), tocilizumab (anti-IL-6 receptor), or canakinumab (anti-IL-1 β), the treatment cannot be terminated at once. In contrast, small molecule inhibitors such as MCC950 (NLRP3 inhibitor) have a much shorter half-life and are rapidly excreted after termination of the therapeutic application.

A balanced treatment of both, pro- and anti-inflammatory responses is probably the most promising therapy option for AP (Figure 2C). The choice of therapeutic targets is crucial to maintain the balance between hyperinflammation and hypoinflammation, as various cytokines directly affect SIRS as well as CARS. One example of this two-sided activity is TNF- α which is instrumental in the establishment of pro-inflammation, but on the other side indirectly and in combination with FasL and TRAIL triggers apoptosis via the TNF receptor signaling pathway (100, 101). A similar phenomenon is observed with IL-1 β , which is decisively involved in the proinflammatory reaction, but simultaneously induces T-cell apoptosis in the intestinal lamina propria, and promotes secondary infections (102). During severe acute pancreatitis the activation of immunosuppressive regulatory T-cells causes a decrease in the number of systemic T-effector cells (31, 40, 47). The experimental depletion of regulatory T-cells in an AP mouse model resulted in a stabilization of the T-effector cell population and prevented infection of necrosis (47). To focus on one specific therapeutic target may be insufficient to rebalance the immune response, therefore parallel approaches should be considered.

Infection of pancreatic necrosis by commensal intestinal bacteria is the most serious complication of acute pancreatitis (24). In the infected necrotic tissue 16S rRNA sequencing detected the presence of facultative pathogenic bacteria, which migrated from the duodenum into the pancreas (47). Recent data show that pathogenic bacteria multiply during acute and chronic pancreatitis (47, 103). It is known that the composition of the intestinal microbiome has a significant influence on the immune response which may implicate a certain therapeutic potential of probiotic food or supplements (104). Animal data have demonstrated that antibiotic iradication of the gut microbiome delays pancreas carcinogenesis by shifting the T-cell response in the tumor (105, 106). The microbiome/inflammation axis could also influence the disease course of acute and chronic pancreatitis (47, 107, 108). In this context the therapeutic administration of antibiotics needs to be re-evaluated and reconsidered.

3 Chronic pancreatitis

Recurrent episodes of pancreatitis, cause morphological changes and the development of functional deficits which lead to the diagnosis of chronic pancreatitis. The clinical pictures of recurrent acute episodes and chronic pancreatitis are very similar, and a relevant percentage of recurrent acute pancreatitis patients develop chronic disease over time. This suggests a general and progressive underlying disease mechanism that needs to be targeted by therapeutic intervention. Chronic pancreatitis is frequently characterized by recurrent episodes of acute inflammation increasing over time the fibrotic replacement of exocrine and endocrine cells (3). In every acute episode the development of a severe course has to be prevented and the intra pancreatic immune homeostasis must be balanced between a pro-inflammatory response and the regenerative and pro-fibrotic reaction.

Progredient fibrosis of the pancreas goes along with the loss of exocrine and endocrine tissue. Over time, more and more acinar cells are destroyed in the acute inflammation phases, and are replaced by fibrotic tissue (Figure 3). The type 2 immune response which is activated in the acute phase in general is responsible for tissue regeneration and wound healing (40). From this point of view the destruction of acinar cells causes a severe, but primarily sterile wound in the pancreatic tissue, which is characterized by a high degree of necrosis and necroptosis (59). As in the case of external wounds, the wound closure is initiated immediately to prevent infection and here the question arises if the fibrotic replacement should be prevented by a specific treatment? On the one hand, fibrosis is part of the regeneration process leading to wound closure. Recent data has shown that fibrosis induced by tissue resident macrophages is essential for the survival of the acute episode of pancreatitis and that fibrosis also plays a protective and regenerative role (41). On the other hand, excessive fibrosis over time can lead to the displacement of exocrine and endocrine tissue and may result in pancreatic insufficiency.

3.1 The pancreatic infiltrate

After necrotic tissue areas in the pancreas have been cleared by phagocytosing macrophages the absence of DAMPs interrupts the inflammatory cascade and the classical activation of macrophages changes to an alternatively activated phenotype (22, 23, 35). Due to the absence of cytokine and chemokine release neutrophil granulocytes are no longer recruited to the pancreas and the resident neutrophils have only a short half-life time and vanish from the organ. Cells of the adaptive immune system are now detectable in high numbers and a distinct population of resident CD4⁺ and CD8⁺ T-cells can be observed (33–35). Beside the infiltration of T-cells also innate lymphoid cells type 2 (ILC2) populate the fibroinflammatory infiltrate (35). The presence and activation status of these immune cells ultimately results in the activation of pancreatic stellate cells and fibroblasts, which drive pancreatic fibrosis and the production of extracellular matrix proteins.

3.2 Alternative activated macrophages activate pancreatic stellate cells

In response to changing microenvironment (LPS, IFNy or cytokines) macrophages can change their phenotype to fulfil



parallel induces the differentiation of alternatively activated macrophages which control the fibrosis/regeneration balance via the release of growth factors like TGF- β and PDGF. Both, pro- and anti-inflammatory signals are involved in the control of the regeneration/fibrosis balance.

different functions. Activated macrophages are usually divided into two categories, M1-like macrophages and M2-like macrophages, with M1 macrophages being mainly involved in pro-inflammatory responses and M2 macrophages (alternatively activated macrophages) being involved in anti-inflammatory responses. Alternatively activated macrophages are the most intensely studied population of immune cells during chronic pancreatitis. M2 macrophages can be further subdivided into different subclasses (M2a, M2b, M2c and M2d) based on the expression patterns of cell surface markers and their release of pro- or anti-inflammatory cytokines (109). During chronic pancreatitis, M2a macrophages, which are characterized by the expression of CD206, ARG1 and FIZZ1, support cell regeneration and fibrosis by releasing TGFβ and IL-10. M2b macrophages, on the other side, are activated via TLR signaling or IL-1 β and mainly support the Th2 response by IL-10 secretion Also, M2c macrophages act immunosuppressive and are responsible for the phagocytosis of apoptotic cells, whereas M2d macrophages release IL-10 and induce vascularization via secretion of VEGF. Alternatively activated macrophages (M2) play an important role in the progressive fibrosis of the pancreas (23) but are also involved in organ regeneration (110). The polarization of

alternatively activated macrophages (M2a) is primarily induced by the Th2 cytokines IL-4/IL-13, which cause phosphorylation and activation of the transcription factor STAT6 via the IL-4/IL-13 receptor. These alternatively activated macrophages secrete various growth factors including TGF-B, and activate pancreatic stellate cells (PSC) as well as acinar cell regeneration (111). Stellate cells are mainly responsible for fibrogenesis during chronic pancreatitis. Following activation by various growth factors like TGF- β and PDGF (112) or cytokines such as IL-6 and TNF- α (113) they secrete extracellular matrix proteins (114). Like leukocytes, stellate cells migrate from the bone marrow into the damaged pancreas (115, 116). Ino et al. were able to show that monocytes, which migrate into the pancreas via the MCP-1/CCR2 signaling pathway, can differentiate into PSCs (117). In turn, PSCs also influence the immune response and secrete cytokines and chemokines such as MCP-1 to further recruit monocytes into the organ (118). Likewise, they can also influence the polarization of macrophages to an alternatively activated phenotype via the secretion of IL-4/IL-13 (23). This immune cell-like behavior shows their critical involvement in the immune response during chronic pancreatitis and the direct connection of tissue fibrosis with the immune

response. The IL-4/IL-13 axis is the most intensively studied signaling pathway affecting pancreatic fibrosis. Therapeutic blockade of the IL-4 receptor (23), or even the use of a depleting IL-4 antibody (35), results in significantly reduced fibrosis of the pancreas. Pascolizumab, an IL-4 depleting antibody is admitted for clinical intervention in moderate to severe asthma (119). Dupilumab, an IL-4/IL-13 receptor blocking antibody, is used in dermatitis as well as asthma (120, 121). Neither of these agents have so far been clinically tested in chronic pancreatitis. One reason for this being the immunosuppressive effect of the therapy, which could increase the risk of developing infected necrosis during an acute episode of pancreatitis. Furthermore, acute pancreatitis was reported as a very rare side effect of treatment with Dupilumab (122).

Macrophage polarization is mainly regulated by T-cells, ILCs, and PSCs. Folias et al. described a significantly reduced number of alternatively activated macrophages in the inflamed pancreas of Rag1-/- mice, which lack T- and B-cells. The defective macrophage polarization resulted in a severely impaired acinar cell regeneration after induction of pancreatitis (111). Thus, alternatively activated macrophages are not only drivers of fibrosis but also essential for tissue regeneration (110). Which makes treatment more complex! This suggests a critical role of Th2-cells for macrophage polarisation into the alternatively activated phenotype. A general regulator of CD4⁺ T-helper cells are regulatory T-cells. T_{regs} have immunosuppressive activity and are particularly increased in the chronically inflamed pancreas (33-35), where they inhibit Th2-cells and ILC2s. The depletion of Tregs resulted in uncontrolled Th2/ ILC2s activation and excessive fibrosis of the pancreas (35). The induction of Tregs could help to prevent excessive fibrosis and one possibility to support the Tregs in their suppressive activity is the administration of IL-2 (123, 124). Interleukin 2, a known T-cell growth factor, can induce and stabilize the Tregs homeostasis (125). However, during the acute phase, excessive activity of Tregs acts also immunosuppressive and can promote the occurrence of infected necrosis (47). Another possibility for a therapeutic intervention with this signaling pathway is the inhibition of IL-4 and IL-13 secretion from their cellular sources. IL-4/IL-13 are primarily secreted by Th2cells or ILC2s, after their migration into the damaged organ. OC000459 is a CRTH2 antagonist that inhibits migration as well as activation of Th2 cells and ILC2s (126). In clinical studies OC000459 was successfully tested in the treatment of asthma (127). Animal data showed that the inhibition of Th2/ILC2 activation reduced the release of IL-4 and IL-13 in the pancreas and inhibited fibrosis (35). The Tcell/macrophage axis just recently came into the focus of pancreatitis research and is still largely not understood. In addition to Th2 cells, also Th22 and Th17 cells are found in the pancreas (33, 34) and seem to have an impact on fibrosis. Beside CD4⁺ T-helper cells, CD8⁺ Tcells are also present in the pancreas (35), but little is known about their function. Experimental studies on liver fibrosis showed that CD8⁺ cells can act fibrolytic by inducing hepatic stellate cell apoptosis (128) or, like shown for dermal fibrosis, secrete IL-4/IL-13 themselves and further enhance fibrosis (129).

3.3 Macrophage independent activation of PSCs

Th22 cells are involved in progressive fibrosis via the secretion of Interleukin 22, which stimulates PSCs to secrete collagen without activating them (130). Likewise, Th17 cells secrete IL-17A which activates PSCs directly via the IL17 receptor and induces the synthesis and secretion of collagen (131). Both cytokines can be depleted by antibodies, which are already approved for the treatment of psoriasis: Fezakinumab, a monoclonal IL-22 antibody (132) and Secukinumab an antibody raised against IL-17A (133). However, IL-22 and IL-17A have also a crucial role in maintaining the intestinal barrier and regulate the secretion of antimicrobial peptides. IL-22 has been demonstrated to be protective in acute pancreatitis, therefore a long-term blockade of these signaling pathways in patients with chronic pancreatitis could have opposing effects, especially in case of an acute pancreatitis episode (134). The currently available data suggest that the treatment of fibrosis in chronic pancreatitis is complex and must consider the involvement of multiple cells of different origin. A general signaling pathway that can be easily manipulated has not yet been identified.

The preventive treatment of fibrosis currently seems not possible without simultaneously affecting cell regeneration as it is the very same growth factors that drive fibrosis as well as regeneration. In addition, also pro-inflammatory mediators such as TNF α and IL-1 β can activate pancreatic stellate cells (112, 113), suggesting that fibrosis is initiated already during the acute phase of pancreatitis. The wound healing process is started during pro-inflammation, directly after the onset of pancreatitis. Consequently there exist many different mechanisms that drive fibrosis, like pro-inflammatory mediators IL-1 β , IL-6, IL17A, and TNF- α , which can activate stellate cells in addition to the other classical growth factors like TGF-B, FGF, and PDGF (35, 114). This makes an anti-fibrotic therapy a challenging task which should be carefully considered. Ultimately, the most effective treatment of fibrosis should be aimed at minimizing the initial pancreatic damage as early as possible to counteract the development of a necrosis-fibrosis sequence at the outset.

4 Conclusion

The most promising therapeutic options for patients suffering from acute and chronic pancreatitis are currently targeted against the involved immune response mechanisms. The inflammatory response is on the other side also a cause of the clinical picture, so a therapeutic approach may address both symptomatic and causal aspects. Previous therapeutic studies which used to target the known triggering pathway of intracellular trypsinogen activation were unsuccessful because these processes are, at the time point of admission to the clinic, normally beyond a point were an intervention could still prevent acinar cell death and the induction of an inflammatory response. Only in a small proportion of patients with confirmed genetic diagnosis a targeted prophylactic therapy which improves protein misfolding or supports intra-acinar protease/anti-protease homeostasis could prevent severe acute episodes and delay disease progression (135).

In all different etiologies of AP it is likewise necessary to prevent or mitigate a severe course of the disease! A strong inflammatory response significantly influences the disease severity and is also responsible for complications like bacterial infected necrosis during the disease course. Targeting a derailing immune response currently appears as the most promising treatment option. Since both proand anti-inflammatory mechanisms (SIRS/CARS) can have detrimental influences on disease severity, beneficial therapies must successfully rebalance this system which requires rapid and controllable working tools.

In a significant proportion of AP-patients we see a progress from recurrent episodes to CP, which underlines the need for early therapeutic intervention. While anti-inflammation is shown to be partially protective in the course of acute pancreatitis, it's profibrotic effects must be considered in recurrent and chronic forms of the disease, as acute and chronic pancreatitis have common underlying disease mechanisms. More experimental studies are needed to unravel the complex immunomodulatory mechanisms involved in the manifestation of the disease in order to uncover further therapeutic options. In parallel, clinical trials should further evaluate the use of already established immune modulating therapeutics.

Author contributions

MS: Conceptualization, Funding acquisition, Writing – original draft. JG: Conceptualization, Funding acquisition, Writing – original draft, Writing – review & editing. SA: Conceptualization, Writing –

References

1. Petrov MS, Yadav D. Global epidemiology and holistic prevention of pancreatitis. Nat Rev Gastroenterol Hepatol (2019) 16:175-84. doi: 10.1038/s41575-018-0087-5

2. Yamamiya A, Tominaga K, Hoshi K, Nagashima K, Minaguchi T, Haruyama Y, et al. The risk factors for progression to chronic pancreatitis in patients with pasthistory of acute pancreatitis: A retrospective analysis based on mechanistic definition. *JCM* (2022) 11:2209. doi: 10.3390/jcm11082209

3. Beyer G, Habtezion A, Werner J, Lerch MM, Mayerle J. Chronic pancreatitis. Lancet (2020) 396:499–512. doi: 10.1016/S0140-6736(20)31318-0

4. Halangk W, Lerch MM, Brandt-Nedelev B, Roth W, Ruthenbuerger M, Reinheckel T, et al. Role of cathepsin B in intracellular trypsinogen activation and the onset of acute pancreatitis. *J Clin Invest* (2000) 106:773-81. doi: 10.1172/JCI9411

5. Halangk W, Lerch MM. Early events in acute pancreatitis. *Gastroenterol Clin North Am* (2004) 33:717-31. doi: 10.1016/j.gtc.2004.07.009

6. Mayerle J, Sendler M, Hegyi E, Beyer G, Lerch MM, Sahin-Tóth M. Genetics, cell biology, and pathophysiology of pancreatitis. *Gastroenterology* (2019) 156:1951–1968.e1. doi: 10.1053/j.gastro.2018.11.081

7. Whitcomb DC, Gorry MC, Preston RA, Furey W, Sossenheimer MJ, Ulrich CD, et al. Hereditary pancreatitis is caused by a mutation in the cationic trypsinogen gene. *Nat Genet* (1996) 14:141–5. doi: 10.1038/ng1096-141

8. Rosendahl J, Witt H, Szmola R, Bhatia E, Ozsvári B, Landt O, et al. (CTRC) variants that diminish activity or secretion are associated with chronic pancreatitis. *Nat Genet* (2008) 40:78–82. doi: 10.1038/ng.2007.44

9. Witt H, Luck W, Hennies HC, Classen M, Kage A, Lass U, et al. Mutations in the gene encoding the serine protease inhibitor, Kazal type 1 are associated with chronic pancreatitis. *Nat Genet* (2000) 25:213–6. doi: 10.1038/76088

original draft. RL: Conceptualization, Writing – original draft. HM: Conceptualization, Writing – original draft. HE: Conceptualization, Writing – original draft. FW: Conceptualization, Writing – original draft, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by Deutsche Forschungsgemeinschaft (DFG SE 2702/2-1, SE 2702/2-3 - 384909139, GRK 2719, GL 1096/1-1 - 508168905).

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10. Witt H, Beer S, Rosendahl J, Chen J-M, Chandak GR, Masamune A, et al. Variants in CPA1 are strongly associated with early onset chronic pancreatitis. *Nat Genet* (2013) 45:1216–20. doi: 10.1038/ng.2730

11. Fjeld K, Weiss FU, Lasher D, Rosendahl J, Chen J-M, Johansson BB, et al. A recombined allele of the lipase gene CEL and its pseudogene CELP confers susceptibility to chronic pancreatitis. *Nat Genet* (2015) 47:518–22. doi: 10.1038/ng.3249

12. Kereszturi É, Szmola R, Kukor Z, Simon P, Ulrich Weiss F, Lerch MM, et al. Hereditary pancreatitis caused by mutation-induced misfolding of human cationic trypsinogen: A novel disease mechanism. *Hum Mutat* (2009) 30:575–82. doi: 10.1002/humu.20853

13. Sharer N, Schwarz M, Malone G, Howarth A, Painter J, Super M, et al. Mutations of the cystic fibrosis gene in patients with chronic pancreatitis. *N Engl J Med* (1998) 339:645–52. doi: 10.1056/NEJM199809033391001

14. Cohn JA, Friedman KJ, Noone PG, Knowles MR, Silverman LM, Jowell PS. Relation between mutations of the cystic fibrosis gene and idiopathic pancreatitis. *N Engl J Med* (1998) 339:653–8. doi: 10.1056/NEJM199809033391002

15. Hegyi P, Rakonczay Z. The role of pancreatic ducts in the pathogenesis of acute pancreatitis. *Pancreatology* (2015) 15:S13–17. doi: 10.1016/j.pan.2015.03.010

16. Romac JM-J, Shahid RA, Swain SM, Vigna SR, Liddle RA. Piezo1 is a mechanically activated ion channel and mediates pressure induced pancreatitis. *Nat Commun* (2018) 9:1715. doi: 10.1038/s41467-018-04194-9

17. Gukovsky I, Gukovskaya AS, Blinman TA, Zaninovic V, Pandol SJ. Early NFkappaB activation is associated with hormone-induced pancreatitis. *Am J Physiol* (1998) 275:G1402-1414. doi: 10.1152/ajpgi.1998.275.6.G1402

18. Sendler M, Weiss F-U, Golchert J, Homuth G, van den Brandt C, Mahajan UM, et al. Cathepsin B-mediated activation of trypsinogen in endocytosing macrophages

increases severity of pancreatitis in mice. *Gastroenterology* (2018) 154:704-718.e10. doi: 10.1053/j.gastro.2017.10.018

19. Zhao Q, Wei Y, Pandol SJ, Li L, Habtezion A. STING signaling promotes inflammation in experimental acute pancreatitis. *Gastroenterology* (2018) 154:1822-1835.e2. doi: 10.1053/j.gastro.2018.01.065

20. Sendler M, Dummer A, Weiss FU, Krüger B, Wartmann T, Scharffetter-Kochanek K, et al. Tumour necrosis factor α secretion induces protease activation and acinar cell necrosis in acute experimental pancreatitis in mice. *Gut* (2013) 62:430–9. doi: 10.1136/gutjnl-2011-300771

21. Gukovskaya AS, Vaquero E, Zaninovic V, Gorelick FS, Lusis AJ, Brennan M-L, et al. Neutrophils and NADPH oxidase mediate intrapancreatic trypsin activation in murine experimental acute pancreatitis. *Gastroenterology* (2002) 122:974–84. doi: 10.1053/gast.2002.32409

22. Wu J, Zhang L, Shi J, He R, Yang W, Habtezion A, et al. Macrophage phenotypic switch orchestrates the inflammation and repair/regeneration following acute pancreatitis injury. *EBioMedicine* (2020) 58:102920. doi: 10.1016/j.ebiom.2020.102920

23. Xue J, Sharma V, Hsieh MH, Chawla A, Murali R, Pandol SJ, et al. Alternatively activated macrophages promote pancreatic fibrosis in chronic pancreatitis. *Nat Commun* (2015) 6:7158. doi: 10.1038/ncomms8158

24. van Dijk SM, Hallensleben NDL, van Santvoort HC, Fockens P, van Goor H, Bruno MJ, et al. Dutch Pancreatitis Study Group. Acute pancreatitis: recent advances through randomised trials. . *Gut* (2017) 66:2024–32. doi: 10.1136/gutjnl-2016-313595

25. Yamamoto Y, Gaynor RB. The rapeutic potential of inhibition of the NF- κ B pathway in the treatment of inflammation and cancer. J Clin Invest (2001) 107:135–42. doi: 10.1172/JCI11914

26. Neuhöfer P, Liang S, Einwächter H, Schwerdtfeger C, Wartmann T, Treiber M, et al. Deletion of IkB α activates RelA to reduce acute pancreatitis in mice through upregulation of Spi2A. *Gastroenterology* (2013) 144:192–201. doi: 10.1053/j.gastro.2012.09.058

27. Algül H, Treiber M, Lesina M, Nakhai H, Saur D, Geisler F, et al. Pancreasspecific RelA/p65 truncation increases susceptibility of acini to inflammationassociated cell death following cerulein pancreatitis. *J Clin Invest* (2007) 117:1490– 501. doi: 10.1172/JCI29882

28. Wartmann T, Mayerle J, Kähne T, Sahin-Tóth M, Ruthenbürger M, Matthias R, et al. Cathepsin L inactivates human trypsinogen, whereas cathepsin L-deletion reduces the severity of pancreatitis in mice. *Gastroenterology* (2010) 138:726–37. doi: 10.1053/j.gastro.2009.10.048

29. Sendler M, Maertin S, John D, Persike M, Weiss FU, Krüger B, et al. Cathepsin B activity initiates apoptosis via digestive protease activation in pancreatic acinar cells and experimental pancreatitis. J Biol Chem (2016) 291:14717–31. doi: 10.1074/jbc.M116.718999

30. Perides G, Weiss ER, Michael ES, Laukkarinen JM, Duffield JS, Steer ML. TNFalpha-dependent regulation of acute pancreatitis severity by Ly-6C(hi) monocytes in mice. J Biol Chem (2011) 286:13327–35. doi: 10.1074/jbc.M111.218388

31. Glaubitz J, Wilden A, van den Brandt C, Weiss FU, Bröker BM, Mayerle J, et al. Experimental pancreatitis is characterized by rapid T cell activation, Th2 differentiation that parallels disease severity, and improvement after CD4+ T cell depletion. *Pancreatology* (2020) 20:1637–47. doi: 10.1016/j.pan.2020.10.044

32. Demols A, Le Moine O, Desalle F, Quertinmont E, Van Laethem JL, Devière J. CD4(+)T cells play an important role in acute experimental pancreatitis in mice. *Gastroenterology* (2000) 118:582–90. doi: 10.1016/s0016-5085(00)70265-4

33. Lee B, Adamska JZ, Namkoong H, Bellin MD, Wilhelm J, Szot GL, et al. Distinct immune characteristics distinguish hereditary and idiopathic chronic pancreatitis. *J Clin Invest* (2020) 130:2705–11. doi: 10.1172/JCI134066

34. Lee B, Namkoong H, Yang Y, Huang H, Heller D, Szot GL, et al. Single-cell sequencing unveils distinct immune microenvironments with CCR6-CCL20 crosstalk in human chronic pancreatitis. *Gut* (2022) 71:1831–42. doi: 10.1136/gutjnl-2021-324546

35. Glaubitz J, Wilden A, Golchert J, Homuth G, Völker U, Bröker BM, et al. In mouse chronic pancreatitis CD25+FOXP3+ regulatory T cells control pancreatic fibrosis by suppression of the type 2 immune response. *Nat Commun* (2022) 13:4502. doi: 10.1038/s41467-022-32195-2

36. Grady T, Liang P, Ernst S, Logsdon C. Chemokine gene expression in rat pancreatic acinar cells is an early event associated with acute pancreatitis. *Gastroenterology* (1997) 113:1966–75. doi: 10.1016/S0016-5085(97)70017-9

37. Yang BM, Demaine AG, Kingsnorth A. Chemokines MCP-1 and RANTES in isolated rat pancreatic acinar cells treated with CCK and ethanol. *In Vitro: Pancreas* (2000) 21:22–31. doi: 10.1097/00006676-200007000-00048

38. Wilden A, Glaubitz J, Otto O, Biedenweg D, Nauck M, Mack M, et al. Mobilization of CD11b+/Ly6chi monocytes causes multi organ dysfunction syndrome in acute pancreatitis. *Front Immunol* (2022) 13:991295. doi: 10.3389/ fmmu.2022.991295

39. Calderon B, Carrero JA, Ferris ST, Sojka DK, Moore L, Epelman S, et al. The pancreas anatomy conditions the origin and properties of resident macrophages. *J Exp Med* (2015) 212:1497–512. doi: 10.1084/jem.20150496

40. Sendler M, van den Brandt C, Glaubitz J, Wilden A, Golchert J, Weiss FU, et al. NLRP3 inflammasome regulates development of systemic inflammatory response and compensatory anti-inflammatory response syndromes in mice with acute pancreatitis. *Gastroenterology* (2020) 158:253–269.e14. doi: 10.1053/j.gastro.2019.09.040

41. Baer JM, Zuo C, Kang L-I, de la Lastra AA, Borcherding NC, Knolhoff BL, et al. Fibrosis induced by resident macrophages has divergent roles in pancreas inflammatory injury and PDAC. *Nat Immunol* (2023) 24(9):1443–57. doi: 10.1038/ s41590-023-01579-x

42. Mittal M, Siddiqui MR, Tran K, Reddy SP, Malik AB. Reactive oxygen species in inflammation and tissue injury. *Antioxidants Redox Signaling* (2014) 20:1126–67. doi: 10.1089/ars.2012.5149

43. Brinkmann V, Reichard U, Goosmann C, Fauler B, Uhlemann Y, Weiss DS, et al. Neutrophil extracellular traps kill bacteria. *Science* (2004) 303:1532–5. doi: 10.1126/ science.1092385

44. Merza M, Hartman H, Rahman M, Hwaiz R, Zhang E, Renström E, et al. Neutrophil extracellular traps induce trypsin activation, inflammation, and tissue damage in mice with severe acute pancreatitis. *Gastroenterology* (2015) 149:1920–1931.e8. doi: 10.1053/j.gastro.2015.08.026

45. Murthy P, Singhi AD, Ross MA, Loughran P, Paragomi P, Papachristou GI, et al. Enhanced neutrophil extracellular trap formation in acute pancreatitis contributes to disease severity and is reduced by chloroquine. *Front Immunol* (2019) 10:28. doi: 10.3389/fimmu.2019.00028

46. Leppkes M, Maueröder C, Hirth S, Nowecki S, Günther C, Billmeier U, et al. Externalized decondensed neutrophil chromatin occludes pancreatic ducts and drives pancreatitis. *Nat Commun* (2016) 7:10973. doi: 10.1038/ncomms10973

47. Glaubitz J, Wilden A, Frost F, Ameling S, Homuth G, Mazloum H, et al. Activated regulatory T-cells promote duodenal bacterial translocation into necrotic areas in severe acute pancreatitis. *Gut* (2023) 72(7):1355–69. doi: 10.1136/gutjnl-2022-327448

48. Esplugues E, Huber S, Gagliani N, Hauser AE, Town T, Wan YY, et al. Control of TH17 cells occurs in the small intestine. *Nature* (2011) 475:514–8. doi: 10.1038/ nature10228

49. Kaplan MJ, Radic M. Neutrophil extracellular traps: double-edged swords of innate immunity. J Immunol (2012) 189:2689–95. doi: 10.4049/jimmunol.1201719

50. Muñoz LE, Boeltz S, Bilyy R, Schauer C, Mahajan A, Widulin N, et al. Neutrophil extracellular traps initiate gallstone formation. *Immunity* (2019) 51:443–450.e4. doi: 10.1016/j.immuni.2019.07.002

51. Weaver CT, Elson CO, Fouser LA, Kolls JK. The th17 pathway and inflammatory diseases of the intestines, lungs, and skin. *Annu Rev Pathol Mech Dis* (2013) 8:477–512. doi: 10.1146/annurev-pathol-011110-130318

52. Shi C, Pamer EG. Monocyte recruitment during infection and inflammation. *Nat Rev Immunol* (2011) 11:762–74. doi: 10.1038/nri3070

53. Ishibashi T, Zhao H, Kawabe K, Oono T, Egashira K, Suzuki K, et al. Blocking of monocyte chemoattractant protein-1 (MCP-1) activity attenuates the severity of acute pancreatitis in rats. J Gastroenterol (2008) 43:79–85. doi: 10.1007/s00535-007-2126-9

54. Bhatia M, Ramnath RD, Chevali L, Guglielmotti A. Treatment with bindarit, a blocker of MCP-1 synthesis, protects mice against acute pancreatitis. *Am J Physiol Gastrointest Liver Physiol* (2005) 288:G1259–1265. doi: 10.1152/ajpgi.00435.2004

55. Thiel FG, Asgarbeik S, Glaubitz J, Wilden A, Lerch MM, Weiss FU, et al. IRAK3mediated suppression of pro-inflammatory MyD88/IRAK signaling affects disease severity in acute pancreatitis. *Sci Rep* (2023) 13:10833. doi: 10.1038/s41598-023-37930-3

56. Fadok VA, Bratton DL, Guthrie L, Henson PM. Differential effects of apoptotic versus lysed cells on macrophage production of cytokines: role of proteases. *J Immunol* (2001) 166:6847–54. doi: 10.4049/jimmunol.166.11.6847

57. Fadok VA, Bratton DL, Konowal A, Freed PW, Westcott JY, Henson PM. Macrophages that have ingested apoptotic cells in *vitro* inhibit proinflammatory cytokine production through autocrine/paracrine mechanisms involving TGF-beta, PGE2, and PAF. *J Clin Invest* (1998) 101:890–8. doi: 10.1172/JCI1112

58. Mareninova OA, Sung K-F, Hong P, Lugea A, Pandol SJ, Gukovsky I, et al. Cell death in pancreatitis: caspases protect from necrotizing pancreatitis. *J Biol Chem* (2006) 281:3370–81. doi: 10.1074/jbc.M511276200

59. Louhimo J, Steer ML, Perides G. Necroptosis is an important severity determinant and potential therapeutic target in experimental severe pancreatitis. *Cell Mol Gastroenterol Hepatol* (2016) 2:519–35. doi: 10.1016/j.jcmgh.2016.04.002

60. He S, Wang L, Miao L, Wang T, Du F, Zhao L, et al. Receptor interacting protein kinase-3 determines cellular necrotic response to TNF-alpha. *Cell* (2009) 137:1100–11. doi: 10.1016/j.cell.2009.05.021

61. Maini R, St Clair EW, Breedveld F, Furst D, Kalden J, Weisman M, et al. Infliximab (chimeric anti-tumour necrosis factor α monoclonal antibody) versus placebo in rheumatoid arthritis patients receiving concomitant methotrexate: a randomised phase III trial. *Lancet* (1999) 354:1932–9. doi: 10.1016/S0140-6736(99)05246-0

62. Fisher CJ, Agosti JM, Opal SM, Lowry SF, Balk RA, Sadoff JC, et al. Treatment of septic shock with the tumor necrosis factor receptor:Fc fusion protein. *N Engl J Med* (1996) 334:1697–702. doi: 10.1056/NEJM199606273342603

63. Colombel J-F, Loftus EV, Tremaine WJ, Egan LJ, Harmsen WS, Schleck CD, et al. The safety profile of infliximab in patients with Crohn's disease: The Mayo Clinic experience in 500 patients. *Gastroenterology* (2004) 126:19–31. doi: 10.1053/j.gastro.2003.10.047

64. Scheller J, Chalaris A, Schmidt-Arras D, Rose-John S. The pro- and antiinflammatory properties of the cytokine interleukin-6. *Biochim Biophys Acta (BBA)* -*Mol Cell Res* (2011) 1813:878–88. doi: 10.1016/j.bbamcr.2011.01.034 65. Zhang R, Shi J, Zhang R, Ni J, Habtezion A, Wang X, et al. Expanded CD14hiCD16- immunosuppressive monocytes predict disease severity in patients with acute pancreatitis. *J Immunol* (2019) 202:2578-84. doi: 10.4049/jimmunol.1801194

66. Rose-John S. IL-6 trans-signaling via the soluble IL-6 receptor: importance for the pro-inflammatory activities of IL-6. *Int J Biol Sci* (2012) 8:1237–47. doi: 10.7150/ ijbs.4989

67. Zhang H, Neuhöfer P, Song L, Rabe B, Lesina M, Kurkowski MU, et al. IL-6 trans-signaling promotes pancreatitis-associated lung injury and lethality. *J Clin Invest* (2013) 123:1019–31. doi: 10.1172/JCI64931

68. Saad MI, Weng T, Lundy J, Gearing LJ, West AC, Harpur CM, et al. Blockade of the protease ADAM17 ameliorates experimental pancreatitis. *Proc Natl Acad Sci U.S.A.* (2022) 119:e2213744119. doi: 10.1073/pnas.2213744119

69. Choy EH, De Benedetti F, Takeuchi T, Hashizume M, John MR, Kishimoto T. Translating IL-6 biology into effective treatments. *Nat Rev Rheumatol* (2020) 16:335–45. doi: 10.1038/s41584-020-0419-z

70. Chen K-L, Lv Z-Y, Yang H-W, Liu Y, Long F-W, Zhou B, et al. Effects of tocilizumab on experimental severe acute pancreatitis and associated acute lung injury. *Crit Care Med* (2016) 44:e664–677. doi: 10.1097/CCM.00000000001639

71. Hoque R, Sohail M, Malik A, Sarwar S, Luo Y, Shah A, et al. TLR9 and the NLRP3 inflammasome link acinar cell death with inflammation in acute pancreatitis. *Gastroenterology* (2011) 141:358–69. doi: 10.1053/j.gastro.2011.03.041

72. Marrache F, Tu SP, Bhagat G, Pendyala S, Osterreicher CH, Gordon S, et al. Overexpression of interleukin-1beta in the murine pancreas results in chronic pancreatitis. *Gastroenterology* (2008) 135:1277–87. doi: 10.1053/j.gastro.2008.06.078

73. Hoque R, Farooq A, Ghani A, Gorelick F, Mehal WZ. Lactate reduces liver and pancreatic injury in Toll-like receptor- and inflammasome-mediated inflammation via GPR81-mediated suppression of innate immunity. *Gastroenterology* (2014) 146:1763–74. doi: 10.1053/j.gastro.2014.03.014

74. Coll RC, Robertson AAB, Chae JJ, Higgins SC, Muñoz-Planillo R, Inserra MC, et al. A small-molecule inhibitor of the NLRP3 inflammasome for the treatment of inflammatory diseases. *Nat Med* (2015) 21:248–55. doi: 10.1038/nm.3806

75. de-Madaria E, Herrera-Marante I, González-Camacho V, Bonjoch L, Quesada-Vázquez N, Almenta-Saavedra I, et al. Fluid resuscitation with lactated Ringer's solution vs normal saline in acute pancreatitis: A triple-blind, randomized, controlled trial. *United Eur Gastroenterol J* (2018) 6:63–72. doi: 10.1177/ 2050640617707864

76. Pan X, Fang X, Wang F, Li H, Niu W, Liang W, et al. Butyrate ameliorates caerulein-induced acute pancreatitis and associated intestinal injury by tissue-specific mechanisms. *Br J Pharmacol* (2019) 176:4446–61. doi: 10.1111/bph.14806

77. Dinarello CA, Simon A, van der Meer JWM. Treating inflammation by blocking interleukin-1 in a broad spectrum of diseases. *Nat Rev Drug Discovery* (2012) 11:633–52. doi: 10.1038/nrd3800

78. Klein AL, Imazio M, Cremer P, Brucato A, Abbate A, Fang F, et al. Phase 3 trial of interleukin-1 trap rilonacept in recurrent pericarditis. *N Engl J Med* (2021) 384:31–41. doi: 10.1056/NEJMoa2027892

79. Dhimolea E. Canakinumab. MAbs (2010) 2:3-13. doi: 10.4161/mabs.2.1.10328

80. De Benedetti F, Gattorno M, Anton J, Ben-Chetrit E, Frenkel J, Hoffman HM, et al. Canakinumab for the treatment of autoinflammatory recurrent fever syndromes. *N Engl J Med* (2018) 378:1908–19. doi: 10.1056/NEJMoa1706314

81. Shakoory B, Carcillo JA, Chatham WW, Amdur RL, Zhao H, Dinarello CA, et al. Interleukin-1 receptor blockade is associated with reduced mortality in sepsis patients with features of macrophage activation syndrome: reanalysis of a prior phase III trial*. *Crit Care Med* (2016) 44:275–81. doi: 10.1097/CCM.000000000001402

82. Leventogiannis K, Kyriazopoulou E, Antonakos N, Kotsaki A, Tsangaris I, Markopoulou D, et al. Toward personalized immunotherapy in sepsis: The PROVIDE randomized clinical trial. *Cell Rep Med* (2022) 3:100817. doi: 10.1016/j.xcrm.2022.100817

83. Kaplan M, Yazgan Y, Tanoglu A, Berber U, Oncu K, Kara M, et al. Effectiveness of interleukin-1 receptor antagonist (Anakinra) on cerulein-induced experimental acute pancreatitis in rats. *Scandinavian J Gastroenterol* (2014) 49:1124–30. doi: 10.3109/00365521.2014.926983

84. Pezzilli R, Billi P, Miniero R, Barakat B. Serum interleukin-10 in human acute pancreatitis. *Dig Dis Sci* (1997) 42:1469–72. doi: 10.1023/a:1018814710291

85. Minshawi F, Lanvermann S, McKenzie E, Jeffery R, Couper K, Papoutsopoulou S, et al. The generation of an engineered interleukin-10 protein with improved stability and biological function. *Front Immunol* (2020) 11:1794. doi: 10.3389/fimmu.2020.01794

86. Palathingal Bava E, George J, Tarique M, Iyer S, Sahay P, Gomez Aguilar B, et al. Pirfenidone increases IL-10 and improves acute pancreatitis in multiple clinically relevant murine models. *JCI Insight* (2022) 7:e141108. doi: 10.1172/jci.insight.141108

87. Rongione A, Kusske A, Kwan K, Ashley S, Reber H, McFadden D. Interleukin 10 reduces the severity of acute pancreatitis in rats. *Gastroenterology* (1997) 112:960–7. doi: 10.1053/gast.1997.v112.pm9041259

88. Devière J, Le Moine O, Van Laethem J-L, Eisendrath P, Ghilain A, Severs N, et al. Interleukin 10 reduces the incidence of pancreatitis after therapeutic endoscopic retrograde cholangiopancreatography. *Gastroenterology* (2001) 120:498–505. doi: 10.1053/gast.2001.21172 89. Garg PK, Singh VP. Organ failure due to systemic injury in acute pancreatitis. Gastroenterology (2019) 156:2008-23. doi: 10.1053/j.gastro.2018.12.041

90. Chaudhry H, Zhou J, Zhong Y, Ali MM, McGuire F, Nagarkatti PS, et al. Role of cytokines as a double-edged sword in sepsis. *In Vivo* (2013) 27:669–84.

91. Hotchkiss RS, Moldawer LL, Opal SM, Reinhart K, Turnbull IR, Vincent J-L. Sepsis and septic shock. *Nat Rev Dis Primers* (2016) 2:16045. doi: 10.1038/nrdp.2016.45

92. Xiao W, Mindrinos MN, Seok J, Cuschieri J, Cuenca AG, Gao H, et al. A genomic storm in critically injured humans. *J Exp Med* (2011) 208:2581–90. doi: 10.1084/ jem.20111354

93. Nascimento DC, Melo PH, Piñeros AR, Ferreira RG, Colón DF, Donate PB, et al. IL-33 contributes to sepsis-induced long-term immunosuppression by expanding the regulatory T cell population. *Nat Commun* (2017) 8:14919. doi: 10.1038/ncomms14919

94. Hotchkiss RS, Monneret G, Payen D. Sepsis-induced immunosuppression: from cellular dysfunctions to immunotherapy. *Nat Rev Immunol* (2013) 13:862-74. doi: 10.1038/nri3552

95. Marik PE, Flemmer M. The immune response to surgery and trauma: Implications for treatment. J Trauma Acute Care Surg (2012) 73:801-8. doi: 10.1097/ TA.0b013e318265cf87

96. Kaukonen K-M, Bailey M, Pilcher D, Cooper DJ, Bellomo R. Systemic inflammatory response syndrome criteria in defining severe sepsis. N Engl J Med (2015) 372:1629–38. doi: 10.1056/NEJMoa1415236

97. Ward NS, Casserly B, Ayala A. The compensatory anti-inflammatory response syndrome (CARS) in critically ill patients. *Clinics Chest Med* (2008) 29:617–25. doi: 10.1016/j.ccm.2008.06.010

98. Schneider CP, Schwacha MG, Chaudry IH. The role of interleukin-10 in the regulation of the systemic inflammatory response following trauma-hemorrhage. *Biochim Biophys Acta (BBA) - Mol Basis Dis* (2004) 1689:22–32. doi: 10.1016/j.bbadis.2004.01.003

99. Venet F, Monneret G. Advances in the understanding and treatment of sepsisinduced immunosuppression. *Nat Rev Nephrol* (2018) 14:121–37. doi: 10.1038/ nrneph.2017.165

100. Hotchkiss RS, Nicholson DW. Apoptosis and caspases regulate death and inflammation in sepsis. Nat Rev Immunol (2006) 6:813-22. doi: 10.1038/nri1943

101. Karki R, Sharma BR, Tuladhar S, Williams EP, Zalduondo L, Samir P, et al. Synergism of TNF-α and IFN- γ Triggers inflammatory cell death, tissue damage, and mortality in SARS-coV-2 infection and cytokine shock syndromes. *Cell* (2021) 184:149–168.e17. doi: 10.1016/j.cell.2020.11.025

102. Roth S, Cao J, Singh V, Tiedt S, Hundeshagen G, Li T, et al. Post-injury immunosuppression and secondary infections are caused by an AIM2 inflammasomedriven signaling cascade. *Immunity* (2021) 54:648–659.e8. doi: 10.1016/ j.immuni.2021.02.004

103. Frost F, Weiss FU, Sendler M, Kacprowski T, Rühlemann M, Bang C, et al. The gut microbiome in patients with chronic pancreatitis is characterized by significant dysbiosis and overgrowth by opportunistic pathogens. *Clin Transl Gastroenterol* (2020) 11:e00232. doi: 10.14309/ctg.0000000000232

104. Akshintala VS, Talukdar R, Singh VK, Goggins M. The gut microbiome in pancreatic disease. *Clin Gastroenterol Hepatol* (2019) 17:290-5. doi: 10.1016/j.cgh.2018.08.045

105. Riquelme E, Zhang Y, Zhang L, Montiel M, Zoltan M, Dong W, et al. Tumor microbiome diversity and composition influence pancreatic cancer outcomes. *Cell* (2019) 178:795–806.e12. doi: 10.1016/j.cell.2019.07.008

106. Pushalkar S, Hundeyin M, Daley D, Zambirinis CP, Kurz E, Mishra A, et al. The pancreatic cancer microbiome promotes oncogenesis by induction of innate and adaptive immune suppression. *Cancer Discovery* (2018) 8:403–16. doi: 10.1158/2159-8290.CD-17-1134

107. Thomas RM, Jobin C. Microbiota in pancreatic health and disease: the next frontier in microbiome research. *Nat Rev Gastroenterol Hepatol* (2020) 17:53–64. doi: 10.1038/s41575-019-0242-7

108. Pagliari D, Saviano A, Newton EE, Serricchio ML, Dal Lago AA, Gasbarrini A, et al. Gut microbiota-immune system crosstalk and pancreatic disorders. *Mediators Inflammation* (2018) 2018:1–13. doi: 10.1155/2018/7946431

109. Yao Y, Xu X-H, Jin L. Macrophage polarization in physiological and pathological pregnancy. *Front Immunol* (2019) 10:792. doi: 10.3389/fimmu.2019.00792

110. Criscimanna A, Coudriet GM, Gittes GK, Piganelli JD, Esni F. Activated macrophages create lineage-specific microenvironments for pancreatic acinar- and β -cell regeneration in mice. *Gastroenterology* (2014) 147:1106–1118.e11. doi: 10.1053/j.gastro.2014.08.008

111. Folias AE, Penaranda C, Su AL, Bluestone JA, Hebrok M. Aberrant innate immune activation following tissue injury impairs pancreatic regeneration. *PloS One* (2014) 9:e102125. doi: 10.1371/journal.pone.0102125

112. Apte MV, Haber PS, Darby SJ, Rodgers SC, McCaughan GW, Korsten MA, et al. Pancreatic stellate cells are activated by proinflammatory cytokines: implications for pancreatic fibrogenesis. *Gut* (1999) 44:534–41. doi: 10.1136/gut.44.4.534

113. Mews P, Phillips P, Fahmy R, Korsten M, Pirola R, Wilson J, et al. Pancreatic stellate cells respond to inflammatory cytokines: potential role in chronic pancreatitis. Gut (2002) 50:535–41. doi: 10.1136/gut.50.4.535

114. Apte M, Pirola RC, Wilson JS. Pancreatic stellate cell: physiologic role, role in fibrosis and cancer. *Curr Opin Gastroenterol* (2015) 31:416–23. doi: 10.1097/MOG.00000000000196

115. Sparmann G, Kruse M-L, Hofmeister-Mielke N, Koczan D, Jaster R, Liebe S, et al. Bone marrow-derived pancreatic stellate cells in rats. *Cell Res* (2010) 20:288–98. doi: 10.1038/cr.2010.10

116. McCarroll JA, Phillips PA, Kumar RK, Park S, Pirola RC, Wilson JS, et al. Pancreatic stellate cell migration: role of the phosphatidylinositol 3-kinase(PI3-kinase) pathway. *Biochem Pharmacol* (2004) 67:1215–25. doi: 10.1016/j.bcp.2003.11.013

117. Ino K, Masuya M, Tawara I, Miyata E, Oda K, Nakamori Y, et al. Monocytes infiltrate the pancreas via the MCP-1/CCR2 pathway and differentiate into stellate cells. *PloS One* (2014) 9:e84889. doi: 10.1371/journal.pone.0084889

118. Wu N, Xu X-F, Xin J-Q, Fan J-W, Wei Y-Y, Peng Q-X, et al. The effects of nuclear factor-kappa B in pancreatic stellate cells on inflammation and fibrosis of chronic pancreatitis. *J Cell Mol Med* (2021) 25:2213–27. doi: 10.1111/jcmm.16213

119. Hart TK, Blackburn MN, Brigham-Burke M, Dede K, Al-Mahdi N, Zia-Amirhosseini P, et al. Preclinical efficacy and safety of pascolizumab (SB 240683): a humanized anti-interleukin-4 antibody with therapeutic potential in asthma. *Clin Exp Immunol* (2002) 130:93–100. doi: 10.1046/j.1365-2249.2002.01973.x

120. Simpson EL, Bieber T, Guttman-Yassky E, Beck LA, Blauvelt A, Cork MJ, et al. Two phase 3 trials of dupilumab versus placebo in atopic dermatitis. *N Engl J Med* (2016) 375:2335–48. doi: 10.1056/NEJMoa1610020

121. Castro M, Corren J, Pavord ID, Maspero J, Wenzel S, Rabe KF, et al. Dupilumab efficacy and safety in moderate-to-severe uncontrolled asthma. *N Engl J Med* (2018) 378:2486–96. doi: 10.1056/NEJMoa1804092

122. Stoner RC, Ophaug S, Simpson E, Hanifin J. Acute pancreatitis in 2 adolescent boys on dupilumab therapy for atopic dermatitis. *Dermatitis* (2021) 32:e21-3. doi: 10.1097/DER.00000000000651

123. Grinberg-Bleyer Y, Baeyens A, You S, Elhage R, Fourcade G, Gregoire S, et al. IL-2 reverses established type 1 diabetes in NOD mice by a local effect on pancreatic regulatory T cells. *J Exp Med* (2010) 207:1871–8. doi: 10.1084/jem.20100209

124. Bluestone JA, Tang Q. T $_{\rm reg}$ cells—the next frontier of cell therapy. Science (2018) 362:154–5. doi: 10.1126/science.aau2688

125. Fontenot JD, Rasmussen JP, Gavin MA, Rudensky AY. A function for interleukin 2 in Foxp3-expressing regulatory T cells. *Nat Immunol* (2005) 6:1142–51. doi: 10.1038/ni1263

126. Pettipher R, Vinall SL, Xue L, Speight G, Townsend ER, Gazi L, et al. Pharmacologic profile of OC000459, a potent, selective, and orally active D prostanoid receptor 2 antagonist that inhibits mast cell-dependent activation of T helper 2 lymphocytes and eosinophils. *J Pharmacol Exp Ther* (2012) 340:473–82. doi: 10.1124/jpet.111.187203

127. Barnes N, Pavord I, Chuchalin A, Bell J, Hunter M, Lewis T, et al. A randomized, double-blind, placebo-controlled study of the CRTH2 antagonist OC000459 in moderate persistent asthma. *Clin Exp Allergy* (2012) 42:38–48. doi: 10.1111/j.1365-2222.2011.03813.x

128. Koda Y, Teratani T, Chu P-S, Hagihara Y, Mikami Y, Harada Y, et al. CD8+ tissue-resident memory T cells promote liver fibrosis resolution by inducing apoptosis of hepatic stellate cells. *Nat Commun* (2021) 12:4474. doi: 10.1038/s41467-021-24734-0

129. Fuschiotti P, Larregina AT, Ho J, Feghali-Bostwick C, Medsger TA. Interleukin-13-producing CD8+ T cells mediate dermal fibrosis in patients with systemic sclerosis. *Arthritis Rheumatism* (2013) 65:236–46. doi: 10.1002/art.37706

130. Xue J, Zhao Q, Sharma V, Nguyen LP, Lee YN, Pham KL, et al. Aryl hydrocarbon receptor ligands in cigarette smoke induce production of interleukin-22 to promote pancreatic fibrosis in models of chronic pancreatitis. *Gastroenterology* (2016) 151:1206–17. doi: 10.1053/j.gastro.2016.09.064

131. Zhao Q, Manohar M, Wei Y, Pandol SJ, Habtezion A. STING signalling protects against chronic pancreatitis by modulating Th17 response. *Gut* (2019) 68:1827–37. doi: 10.1136/gutjnl-2018-317098

132. Guttman-Yassky E, Brunner PM, Neumann AU, Khattri S, Pavel AB, Malik K, et al. Efficacy and safety of fezakinumab (an IL-22 monoclonal antibody) in adults with moderate-to-severe atopic dermatitis inadequately controlled by conventional treatments: A randomized, double-blind, phase 2a trial. *J Am Acad Dermatol* (2018) 78:872–881.e6. doi: 10.1016/j.jaad.2018.01.016

133. Langley RG, Elewski BE, Lebwohl M, Reich K, Griffiths CEM, Papp K, et al. Secukinumab in plaque psoriasis — Results of two phase 3 trials. *N Engl J Med* (2014) 371:326–38. doi: 10.1056/NEJMoa1314258

134. Xue J, Nguyen DTC, Habtezion A. Aryl hydrocarbon receptor regulates pancreatic IL-22 production and protects mice from acute pancreatitis. *Gastroenterology* (2012) 143:1670-80. doi: 10.1053/j.gastro.2012.08.051

135. Pesei ZG, Jancsó Z, Demcsák A, Németh BC, Vajda S, Sahin-Tóth M. Preclinical testing of dabigatran in trypsin-dependent pancreatitis. *JCI Insight* (2022) 7:e161145. doi: 10.1172/jci.insight.161145